

Guidelines for management of uveal melanoma (2025)

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ABSTRACT

The guidelines for uveal melanoma have undergone substantial transformations with the introduction of advanced diagnostic and therapeutic technologies. This article aims to elucidate the complex application of contemporary advancements in the management of uveal melanoma, a field that holds significant promise yet presents unique challenges due to its intricate biological behavior. Historically, the therapeutic landscape of uveal melanoma was predominantly characterized by surgical interventions. However, advancements in medical science, particularly the development of immunotherapies and targeted therapies, have shifted this paradigm. When combined with the rapid expansion of genomic and proteomic data, the role of these novel treatments in managing uveal melanoma has been firmly established, transitioning from experimental stages to clinical practice. Acknowledging this transformative potential, collaborations between the Ophthalmic Oncology Subcommittee of the International Ophthalmological Society and the Melanoma Research Consortium have been instrumental. These distinguished organizations convened a panel of experts to examine and integrate the latest advancements from both national and international sources. Their scope extended beyond the implementation of innovative therapies for uveal melanoma, encompassing the identification of existing barriers and the outlining of future directions. Following thorough deliberations, the consensus synthesized in this document serves as a guiding framework for ophthalmologists and oncologists, facilitating the effective incorporation of contemporary advancements into treatment strategies and the advancement of clinical research. This guideline aims to provide a comprehensive framework, ensuring that clinical decisions are not only informed but transformed by the latest scientific developments. By building upon existing knowledge while adhering to the highest standards of originality, this document exemplifies both innovation and scholarly integrity, aligning with the principles of esteemed journals such as *Ophthalmology* and *Cancer Research*.

INTRODUCTION

Uveal melanoma (UM) is the most common primary malignant intraocular tumor in adults, significantly threatening the life and visual function of affected individuals. It constitutes approximately 79% to 81% of ocular melanomas and 3% to 5% of all melanoma cases [1]. UM originates from melanocytes located within the uveal tract, which includes the iris, ciliary body, and choroid. Approximately 90% of UM cases occur in the choroid,

while the remaining 10% are found in the iris and ciliary body, typically presenting as monocular [2]. The global incidence rate of uveal melanoma ranges from 0.0001% to 0.0009% [3], with significant geographic and racial disparities. The highest prevalence is observed among Caucasians, followed by Asians, with a lower incidence in individuals of African descent. Additionally, the prevalence

is slightly higher in males compared to females. Approximately 50% of patients with UM eventually develop hematogenous metastasis, predominantly affecting the liver. A study conducted at Wills Eye Hospital in the United States reported that the 5-, 10-, and 15-year metastasis rates for large UM tumors were 35%, 49%, and 67%, respectively, while the rates for medium-sized tumors were 14%, 26%, and 37% [4, 5]. A long-term follow-up study of 1,553 Chinese UM patients at Beijing Tongren Hospital of Capital Medical University revealed 5-, 10-, and 15-year metastasis rates of 19%, 27%, and 31%, respectively. Additionally, the Eye, Ear, Nose and Throat Hospital of Fudan University documented 5- and 10-year metastasis rates of 20% and 30% for UM, respectively [6]. The age at initial diagnosis for most international patients ranges from 50 to 70 years [7], whereas Chinese patients are typically diagnosed at a younger age, with an average of approximately 45 years [8].

In recent years, advancements in diagnostic and therapeutic methodologies have significantly improved the local control rate of UM. Nevertheless, these advancements have not translated into a corresponding increase in

PATHOGENESIS

Despite the high metastatic potential of UM, metastatic lesions are detectable in fewer than 4% of patients at the time of initial diagnosis [10]. It is likely that most patients have clinically undetectable micrometastases at diagnosis, which supports the classification of UM as a systemic disease [11]. The pathogenesis of UM is intricately linked to molecular genetics, epigenetics, cellular immunity, and environmental factors. Chromosomal abnormalities in UM cells include deletions of chromosome 3 and amplifications of the long arms of chromosomes 6 and 8. Monosomy of chromosome 3 is the most common karyotypic aberration, occurring in approximately 50% to 60% of patients [12]. The deletion of chromosome 3 is a prognostic indicator for the increased likelihood of metastasis in UM [13]. Additionally, chromosome 3 deletion is associated with a higher propensity for metastasis and a poor prognosis [13]. In contrast, amplification of the long arm of chromosome 6 is frequently observed in patients with low-risk UM, whereas amplification of chromosome 8 correlates with a poor prognosis [14].

The primary oncogenic mutations in UM include

survival rates, as UM continues to exhibit high rates of metastasis, morbidity, and mortality [1,4,8]. The liver (89%), lungs (29%), and bones (17%) are the most common sites for metastasis [1]. Importantly, the median survival time for patients with metastatic UM is only 6 to 12 months [9].

To standardize the clinical diagnosis and treatment of UM, the Ophthalmology Committee of the China Medical Education Association, the Oculoplastic and Orbital Disease Group of the Ophthalmology Branch of the Chinese Medical Association, and the Ocular Tumor Committee of the Chinese Anti-Cancer Association convened a panel of domestic experts specializing in fundoplication, ocular tumors, ocular pathology, radiation therapy, and interventional therapy. These experts engaged in comprehensive discussions and developed a consensus statement on the diagnosis and treatment of UM. This consensus, informed by both domestic and international literature and tailored to the specific context of the country, serves as a reference for clinicians in ophthalmology and related fields.

gain-of-function mutations in GNAQ or GNA11 [15], deletion of the oncogene histone deubiquitylating enzyme BAP1 [16], and mutation of the splicing factor SF3B1 gene [17]. Notably, gain-of-function mutations in GNAQ and GNA11 are present in 91% of UM patients and are considered critical driver mutations. These mutations activate multiple signaling pathways, including mitogen-activated protein kinase kinase, protein kinase C, and Yes-associated proteins, thereby promoting the malignant transformation of UM [18]. The histone deubiquitinating enzyme BAP1 is a critical oncogene in the pathogenesis of UM, with its deletion occurring in 50% of UM patients. The loss of BAP1 function impairs UM cell adhesion, triggers cytoskeletal remodeling, and facilitates the metastasis of UM cells [19]. Additionally, mutations in the splicing factor SF3B1 gene are detected in approximately 25% of UM patients, leading to the production of aberrant transcription products within the tumor tissue. These abnormal transcription products significantly contribute to tumor proliferation in UM [17].

It is significant to note that certain patients with UM

do not exhibit the typical genetic mutations commonly associated with the disease, yet they experience some of the poorest prognoses. This observation suggests the involvement of additional factors in UM pathogenesis. Recent studies have demonstrated that epigenetic imbalances play a crucial role in the development and progression of UM [20]. For instance, histone H3K27 methylation in UM can inhibit immune antigen presentation, thereby facilitating immune escape [21]. Furthermore, abnormal chromosomal conformations in UM can create oncogenic enhancers, activating the expression

DIAGNOSIS

Symptoms and signs

UM encompasses melanomas located in the iris, ciliary body, and choroid, *e.* Choroidal melanoma typically manifests with symptoms such as persistent photopsia, scotomas, visual field defects, or vision loss, although some patients may remain asymptomatic. Choroidal melanomas are classified into three morphological types: dome-shaped, mushroom-shaped, and flat diffuse. Dome-shaped choroidal melanomas are restricted to the choroidal stroma and have not breached Bruch's membrane. In contrast, mushroom-shaped choroidal melanomas are bilobed tumors that penetrate Bruch's membrane and extend into the subretinal space. Flat diffuse melanomas exhibit a flat, widespread growth pattern and are often misdiagnosed as choroidal nevi. Choroidal melanoma is characterized by the absence of retinal trophoblastic vessels and frequently leads to exudative retinal detachment, occasionally accompanied by vitreous hemorrhage, which results in visual impairment [29].

Iris melanoma is clinically uncommon and is often discovered incidentally, presenting with changes in iris pigmentation (heterochromia) and pupil distortion [30]. Iris melanomas are categorized into nodular and diffuse types,

Ophthalmologic examination

Patients necessitate a thorough evaluation of visual acuity, intraocular pressure, the anterior segment of the eye, and the fundus. The primary ophthalmologic examination techniques include slit lamp microscopy and indirect ophthalmoscopy, with occasional use of gonioscopy or

of the oncogene NTS and promoting malignant transformation [22]. Additional epigenetic factors, including m6A RNA methylation, histone lactylation, and long non-coding RNAs, are also implicated in UM pathogenesis [23-25]. Importantly, pharmacological interventions targeting these epigenetic abnormalities, such as DNA methyltransferase inhibitors (DNMTi) and histone deacetylase inhibitors (HDACi), have been shown to effectively inhibit the malignant transformation of UM [26-28].

manifesting as pigmented, non-pigmented, or mixed pigmented tumors. Nodular iris melanomas are characterized by a dome-shaped morphology and the presence of dilated trophoblastic vessels. In contrast, diffuse iris melanomas are marked by increased pigmentation of the iris without the formation of a discernible mass. As the condition advances, iris melanoma can lead to complications such as iris ectasia, anterior chamber hemorrhage, and secondary glaucoma. Further progression may result in tumor seeding within the anterior chamber and potential extraocular dissemination.

Ciliary body melanoma is often obscured by the iris, resulting in minimal early-stage symptoms. Consequently, it is frequently not diagnosed until more pronounced symptoms, such as lens displacement, retinal detachment, or extra-scleral extension, become apparent. Upon pupil dilation, the mass may be visualized posterior to the lens. Additionally, the presence of outer scleral trophoblastic blood vessels, known as anterior sentinel vessels, serves as a significant indicator for the potential presence of melanoma [31].

transillumination. All patients underwent assessment using a slit lamp microscope for the anterior segment and an indirect ophthalmoscope for the posterior segment to ascertain the location, shape, pigmentation, vascular distribution, tumor margin morphology, distance from the

macula and optic disc, as well as involvement of the ciliary body and cornea, anterior extrascleral extension, and to identify secondary pathologies such as malignant transformation of a choroidal nevus. This includes anterior sentinel vessels on the scleral surface, cataracts, subretinal

Imaging examination

Photography of the anterior segment of the eye and fundus serves as an objective method to document the distribution, morphology, pigmentation, and presence of trophoblastic vessels in iris and ciliary body tumors, as well as extra-scleral extensions, anterior sentinel vessels of the sclera, and other lesions. It also captures the location and morphology of the lens and pupil. In cases of ciliary body and choroidal melanomas, it is essential to achieve full pupil dilation for thorough examination. Simultaneous recording of the peripheral fundus is crucial to delineate the tumor's location and extent, as well as any associated exudative retinal detachment. Ultra-wide-angle fundus imaging provides a more comprehensive depiction of the choroidal melanoma's size and extent, and its relationship with the macula and optic disc. This imaging modality is particularly advantageous for visualizing multiple scattered choroidal melanomas concurrently and for assessing the extent and severity of combined exudative retinal detachment in relation to the tumor. In contrast, conventional fundus photography offers a clearer and more objective representation of the tumor surface details, including color and pigment distribution. For the assessment of choroidal melanoma, a combination of diagnostic techniques is recommended [32].

Ultrasonography and ultrasound biomicroscopy are pivotal in this regard. Ultrasonography, in particular, is the most prevalent method for determining the volume of choroidal melanoma and plays a vital role in its screening, diagnosis, treatment, and monitoring. On ocular ultrasound A-scan, choroidal melanoma is characterized by moderately low internal reflectivity, with a peak at the tumor's apex and a subsequent gradual decline in reflectivity as the sound waves traverse the mass. Typically, on ultrasound imaging, choroidal melanoma presents as dome-shaped, mushroom-shaped, or flat, often exhibiting a positive scooping sign and involving a choroidal depression. Ultrasonography is instrumental in determining whether a tumor has invaded the orbit. The application of

fluid, or tumors with orange pigment. Gonioscopy is employed to detect iris or ciliary body melanoma affecting the anterior chamber angle, while transillumination, either transscleral or pupillary, is utilized to determine the extent of ciliary body involvement.

color Doppler flow ultrasound facilitates the differentiation between a solid tumor and a hemorrhage by assessing the blood supply and flow within the lesion. In patients with choroidal melanoma, ultrasonography, when employed alongside conventional methods, allows for dynamic observation of blood perfusion within the tumor. Through post-processing and image analysis, perfusion curves and quantitative diagnostic parameters can be derived [33]. Choroidal melanoma typically exhibits the circulatory metabolic characteristics of malignant tumors, with the ultrasonography time-intensity curve demonstrating a rapid-in-rapid-out pattern [34]. Ultrasound bio-microscopy is frequently utilized to evaluate tumors of the iris and ciliary body. Its high resolution enables detailed visualization of the lesion's internal structure and its potential invasion into surrounding tissues, which is crucial for assessing the volume of ciliary body melanoma, the presence of extrascleral spread, and the extent of iris tumor invasion into the ciliary body (Figure 1).

Orbital magnetic resonance imaging (MRI) offers high resolution and is particularly effective in detecting the spread of extrascleral tumors. It is considered more valuable than computed tomography (CT) in the imaging of UM and its relationship to the orbit, especially in cases involving large tumors. Typical MRI indicators of choroidal melanoma include a mass located posterior to the equator, often presenting as thick, mushroom-shaped, or hemispherical. However, the diagnostic value of the mass's morphology is limited. Characteristically, MRI of UM exhibits a high signal intensity on T1-weighted images (T1WI) and a low signal intensity on T2-weighted images (T2WI), relative to the signal of the brain's gray matter. The majority of UMs demonstrate mild enhancement on contrast-enhanced scans, in contrast to melanocytomas, which do not enhance, and most other tumors, which show moderate to significant enhancement [35]. Clinically, retinal detachment is frequently observed. It is recommended to use the brain's gray matter as a reference

for determining the signal intensity of ocular masses, as it provides higher diagnostic accuracy compared to using the vitreous body as a reference. Although orbital CT is less commonly employed for the diagnostic imaging of UM, it retains value in imaging large tumors and assessing orbital invasion (Figure 2).

Fluorescein fundus angiography (FFA) and indocyanine green angiography (ICGA) are instrumental in the evaluation of choroidal melanoma. In the early stages, FFA typically reveals patchy hyperfluorescence, while in the later stages, diffuse fluorescence leakage may be observed due to continuous leakage from the tumor vasculature. Prior to the tumor breaching Bruch's membrane, ICGA images do not display the choroidal blood vessels within the tumor, attributable to either dense pigmentation or an absence of blood vessels within the tumor. However, once the tumor penetrates Bruch's membrane, ICGA examination reveals retinal macrovessels at the tumor site with varying diameters and disorganized alignments, alongside various abnormal vascular formations within the tumor. Post-radiation therapy, tumors may exhibit pre-retinal or subretinal neovascularization, indicative of radiation retinopathy as a complication of radiation treatment [36].

Optical Coherence Tomography (OCT) is a valuable imaging modality for detecting subtle retinal abnormalities,

such as subretinal fluid, intraretinal edema, irregular morphology of the photoreceptor layer, and tumor-induced compression of the choroid. It also facilitates the identification of choroidal defects within tumors, as well as the cross-sectional configuration of choroidal capillaries, orange pigment, and choroidal lesions compressed by tumors. Compared to ultrasonography, OCT offers superior advantages in measuring the thickness of small choroidal melanomas [37]. OCT of the anterior segment of the eye is recommended for examining iris melanomas; however, the basal margin of the tumor may be obscured by pigmentation. Additionally, fundus coherence photomicrographic angiography can be employed to monitor macular microangiopathy following radiation therapy, thereby providing a basis for clinical treatment [38].

Positron emission tomography-computed tomography (PET-CT) scans exhibit high sensitivity and predictive value in monitoring systemic metastases in patients with UM. For individuals with UM metastasis, PET-CT is recommended for the early detection of metastases and accurate tumor staging, which is crucial for effective patient management and follow-up. Furthermore, PET-CT is instrumental in identifying primary foci of choroidal metastatic carcinoma [39].

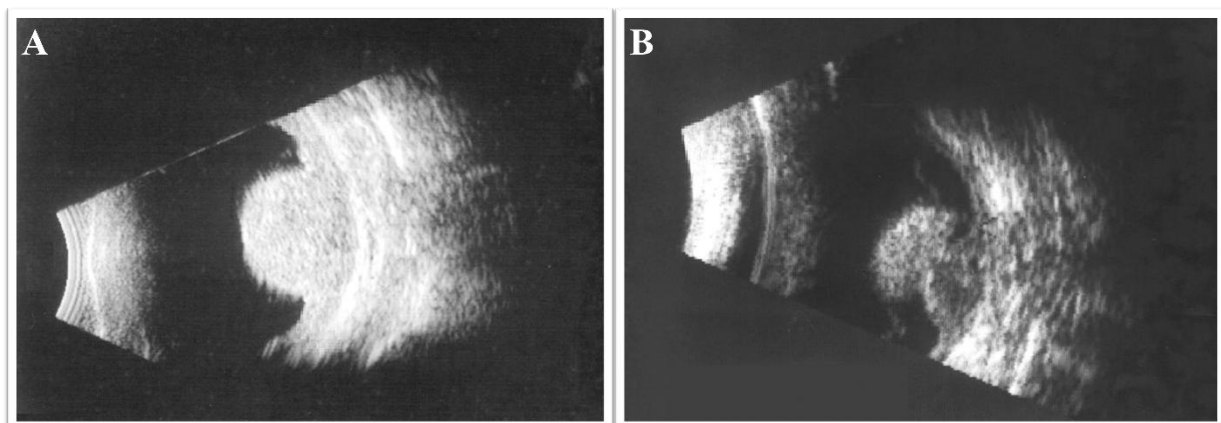


Figure 1: B-scan Ultrasound Images of Choroidal Melanoma. A) The choroidal melanoma demonstrates a mushroom-shaped growth. B) The tumor shows an excavation sign.

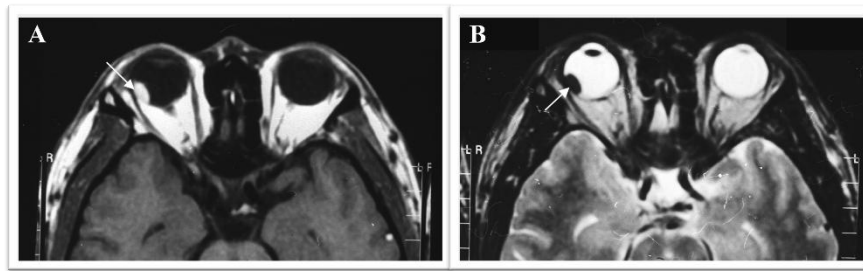


Figure 2: Axial Orbital MRI Images of Choroidal Melanoma. A) High signal on T1WI (arrow). B) Low signal on T2WI (arrow).

Diagnosis and differential diagnosis

A range of lesions present with clinical features similar to those of choroidal melanoma. Common differential diagnoses include optic disc melanocytoma, choroidal melanocytosis, choroidal metastatic carcinoma, retinal pigment epithelial adenoma or adenocarcinoma, choroidal nevus, peripheral exudative hemorrhagic choroidal retinopathy, congenital hypertrophy of the retinal pigment epithelium, hemorrhagic detachment of the retina or pigmented epithelium, choroidal hemangioma, polypoid choroidal lesions, and age-related macular degeneration.

In cases where patients exhibit atypical clinical presentations, diagnostic vitreous surgery and fine-needle aspiration biopsy (FNAB) can provide clarity in diagnosis [40]. For choroidal melanoma biopsy sampling, 25-gauge or 27-gauge diagnostic vitreous surgery is recommended,

as it minimizes the risk of hemorrhage and retinal detachment while ensuring adequate sample collection under visual guidance. FNAB, performed via the scleral or transvitreal route, can preserve retinal integrity when executed correctly, with a low incidence of ocular complications. Conducting FNAB prior to initiating therapeutic interventions can enhance treatment safety [41]. Tumor samples obtained through FNAB are valuable for molecular pathology testing, which aids in prognostic assessment in addition to confirming the diagnosis. Nevertheless, FNAB is associated with potential risks, including insufficient sampling, medical injury, and the extraocular spread of tumors; therefore, it should be employed with caution [42].

International staging

UM is primarily staged using two principal systems, both of which are predicated on tumor thickness and basal maximum diameter. The first of these systems was developed by the Collaborative Ocular Melanoma Study (COMS) Group [43], while the second, known as the TNM staging system, was introduced by the American Joint Committee on Cancer (AJCC) in 1968 [44]. In the COMS staging system, tumors are categorized as small (thickness between 1.5 and 2.4 mm and maximum basal diameter between 5 and 16 mm), medium (thickness between 2.5 and 10.0 mm and maximum basal diameter ≤ 16 mm), and large (thickness >10.0 mm or maximum basal diameter >16 mm), based on tumor thickness and maximum basal diameter. Conversely, the TNM staging system classifies tumors into stages T1, T2, T3, and T4, where "T" refers to the characteristics of the primary tumor,

including its volume and infiltration into surrounding tissues; "N" represents the degree and extent of regional lymph node involvement; and "M" denotes the presence of distant metastasis. There is a partial overlap between small and medium-sized tumors in the COMS staging system and stages T1 and T2 in the TNM staging system, as well as between large tumors in COMS staging and stages T3 and T4 in TNM staging.

In the 8th edition of the TNM staging system, introduced by the AJCC in 2018, a more detailed staging of iris melanomas and the extra-scleral spread of tumors was implemented. Primary tumors are categorized into stages T1 to T4 based on clinical characteristics: stage T1 includes tumors confined to the iris; stage T2 encompasses tumors invading the ciliary body and/or choroid; stage T3 involves tumors invading the ciliary body and/or choroid

with scleral infiltration; and stage T4 includes tumors with extrascleral spread. The 8th edition of the TNM staging system also updated the staging criteria for ciliary body and choroidal melanoma (Table 1).

UM is classified into stage N0, indicating no lymph node involvement, and stage N1, indicating lymph node involvement. Additionally, it is categorized into stage M0,

indicating no metastasis, and stage M1, indicating metastasis. The metastatic lesions are further classified into stages M1a to M1c based on their size.

The AJCC evaluates patient prognosis using the TNM staging system, categorizing patients into seven distinct groups based on the assessed prognosis (Table 2).

Table 1. TNM Staging of Primary Tumor Characteristics in Ciliary Body and Choroidal Melanoma, 8th Edition of AJCC.

Primary Tumor Characteristics and Stages (T)		Tumor Characteristics
T1		1. Tumor base diameter between 3.0~9.0 mm, thickness ≤ 6 mm
		2. Tumor base diameter between 9.1~12.0 mm, thickness ≤ 3 mm
	T1a	T1 tumor without involvement of the ciliary body, no extrascleral extension
	T1b	T1 tumor involving the ciliary body
	T1c	T1 tumor without involvement of the ciliary body, extrascleral extension with maximum diameter ≤ 5 mm
T2	T1d	T1 tumor involving the ciliary body, extrascleral extension with maximum diameter ≤ 5 mm
		1. Tumor base diameter < 9.0 mm, thickness between 6.1~9.0 mm
		2. Tumor base diameter between 9.1~12.0 mm, thickness between 3.1~9.0 mm
		3. Tumor base diameter between 12.1~15.0 mm, thickness ≤ 6.0 mm
		4. Tumor base diameter between 15.1~18.0 mm, thickness ≤ 3.0 mm
T3	T2a	T2 tumor without involvement of the ciliary body, no extrascleral extension
	T2b	T2 tumor involving the ciliary body
	T2c	T2 tumor without involvement of the ciliary body, extrascleral extension with maximum diameter ≤ 5 mm
	T2d	T2 tumor involving the ciliary body, extrascleral extension with maximum diameter ≤ 5 mm
		1. Tumor base diameter between 3.1~9.0 mm, thickness between 9.1~12.0 mm
T4		2. Tumor base diameter between 12.1~15.0 mm, thickness between 6.1~15.0 mm
		3. Tumor base diameter between 15.1~18.0 mm, thickness between 3.1~12.0 mm
	T3a	T3 tumor without involvement of the ciliary body, no extrascleral extension
	T3b	T3 tumor involving the ciliary body
	T3c	T3 tumor without involvement of the ciliary body, extrascleral extension with maximum diameter ≤ 5 mm
	T3d	T3 tumor involving the ciliary body, extrascleral extension with maximum diameter ≤ 5 mm
		1. Tumor base diameter between 12.1~15.0 mm, thickness > 15.0 mm
		2. Tumor base diameter between 15.1~18.0 mm, thickness > 12.1 mm

	3. Tumor base diameter >18.0 mm, thickness not limited
T4a	T4 tumor without involvement of the ciliary body, no extrascleral extension
T4b	T4 tumor involving the ciliary body
T4c	T4 tumor without involvement of the ciliary body, extrascleral extension with maximum diameter ≤5 mm
T4d	T4 tumor involving the ciliary body, extrascleral extension with maximum diameter ≤5 mm
T4e	Any tumor size with extrascleral extension with maximum diameter >5 mm

Table 2. TNM Staging Characteristics for Prognostic Classification of Ciliary Body and Choroidal Melanoma by the American Joint Committee on Cancer.

Prognostic Classification	Primary Tumor Characteristics (T)	Lymph Node Involvement (N)	Distant Metastasis (M)
I	T1a	N0	M0
II	II A T1b~T1d, T2a	N0	M0
	II B T2b, T3a	N0	M0
III	IIIA T2c, T2d, T3b, T3c, T4a	N0	M0
	IIIB T3d, T4b, T4c	N0	M0
	IIIC T4d, T4e	N0	M0
IV	Any T	N1	M1a-M1c

TREATMENT

In recent years, significant advancements in the treatment of UM have been achieved, characterized by rapid and multifaceted developments. Commonly utilized therapies include peripapillary radiation therapy and enucleation. Additionally, a variety of eye-preserving treatments are available, such as transpupillary thermotherapy (TTT), particle radiation therapy, stereotactic radiation therapy, and local excision of ocular tumors. The selection of an appropriate UM treatment regimen is contingent upon several factors, including tumor size, location, and associated features like retinal detachment, vitreous hemorrhage, or retinal infiltration (Table 1). Furthermore, the choice of treatment modality must take into account the tumor's volume and location, as well as its associated characteristics. It is also essential to consider the patient's age, overall health, condition of the fellow eye, and personal preferences (Table 3). Principles for developing treatment recommendation levels [45] involve a simplified scoring system designed to distinguish and indicate both the strength of recommendations and the quality of

evidence underlying those recommendations. The strength of recommendation is classified into two categories: strong recommendation [I] and cautious recommendation [II]. A strong recommendation [I] is attributed to interventions where the benefits substantially outweigh the adverse effects. In contrast, a cautious recommendation [II] is assigned to interventions where the evidence is either of low quality or poorly directed, resulting in an insufficient assessment of benefits versus risks. The quality of evidence is further categorized into four levels.

Level A (high quality), where the evidence is robust and the probability of future research significantly altering the current conclusions is low.

Level B (intermediate quality), where the evidence is moderate and there is a possibility that further research could impact the current conclusions.

Level C (low quality), where the evidence is inadequate and the likelihood of future research significantly affecting the current conclusions is high.

Level D (very low quality): Current consensus among

experts is limited, or there is a paucity of evidence at this juncture.

Table 3. Treatment Methods for UM.

Treatment Method	Indication	Treatment Outcome	Complications	Supplementary Information	Recommendation Grade
Radiotherapy					
Epiretinal brachytherapy (^{125}I , ^{106}Ru)	Small, medium, and large UM with a basal diameter <20 mm	Good local tumor control	Vision loss; tumor recurrence	Adjust dose to delay vision loss	I , A
Plaque brachytherapy	Medium and large UM	Good local tumor control	Vision loss; neovascular glaucoma; tumor recurrence	Not available in all ophthalmic institutions	II , B
Stereotactic radiotherapy	UM near the optic disc; unsuitable for brachytherapy or surgery	Good local tumor control	Vision loss; radiation-related complications; tumor recurrence	Not available in all ophthalmic institutions	II , C
Laser treatment					
TTT	Small UM; local recurrence of UM; adjuvant therapy	Improved local tumor control	Vision loss; extraocular tumor recurrence	Occasionally used for small UM nasal to the optic disc	I , A
Photodynamic therapy (PDT)	Small UM	Uncertain	Tumor recurrence	Avoids radiotherapy complications; not widely used	II , B
Surgery					
Enucleation or combined with brachytherapy	Medium and large UM with a small basal diameter	Uncertain	Retinal detachment; vision loss; enucleation; tumor recurrence	Rarely combined with brachytherapy; combined to reduce recurrence risk	II , B
Intraocular resection or combined with radiotherapy	Medium and large UM; tumor toxicity syndrome	Inconsistent results	Transient intraocular hemorrhage; rare tumor dissemination	Only adopted in some ophthalmic institutions	II , B
Enucleation	Large UM; with neovascular glaucoma or extensive retinal detachment	100% local tumor control if completely resected	Socket-related complications; orbital tumor recurrence	Good cosmetic effects with orbital implants and artificial eyes	I , A

Exenteration	Extraocular extension of UM	100% local tumor control if completely resected	Orbital tumor recurrence	Rarely used	II, B
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Abbreviation: UM, uveal Melanoma; TTT, transpupillary thermotherapy.

Patch radiation therapy

Patch radiation therapy, a modality of brachytherapy, is considered the preferred treatment for small to medium-sized tumors, defined as those with a maximum tumor base diameter of ≤ 18 mm and a thickness of ≤ 10 mm [46]. Commonly utilized radioisotopes in this treatment include iodine-125 (^{125}I) and ruthenium-106 (^{106}Ru). This technique involves the precise placement of a curved dressing containing radioactive particles on the scleral surface corresponding to the tumor location, allowing radiation to penetrate through the sclera and target the tumor. It is essential that the radiation dose delivered to the tumor apex achieves a range of 80-100 Gy. This

therapeutic approach has been shown to effectively control tumor growth while preserving ocular integrity and partial visual function. According to the COMS staging, the five-year survival rate for patients with intermediate-sized tumors following patch radiation therapy (82%) is comparable to that of enucleation alone (81%) [47]. However, the primary complications associated with patch radiation therapy include local radiation-induced neovascular glaucoma, radiation retinopathy, and radiation optic neuropathy, in addition to intraoperative hemorrhage, adjacent tissue injury, and postoperative infection related to the surgical procedure.

TTT

TTT is a non-invasive therapeutic modality that employs an 810 nm infrared diode laser to elevate the temperature of choroidal tumors to between 45 and 60°C. This thermal increase induces vascular occlusion and subsequent tumor necrosis. TTT can penetrate to a maximum depth of 4 mm, making it particularly suitable for small, thick tumors located at the optic disc or outside the macula, especially in cases of primary tumors or local recurrences following complete tumor excision via vitrectomy. It demonstrates enhanced efficacy in treating choroidal melanomas of significant thickness. Tumors exceeding 3 mm in thickness are recommended to be treated with a combination of

proton beam radiotherapy (PBRT) and TTT, commonly referred to as "sandwich" therapy. The advantages of TTT include precise laser targeting, immediate induction of tumor necrosis, minimal damage to adjacent healthy choroidal tissue, ease of administration, outpatient treatment feasibility, and the potential for repeated applications. However, TTT is associated with a propensity for tumor recurrence and potential complications, including anterior retinal membrane formation, branch retinal vein occlusion, retinal traction, and secondary retinal detachment at the foramen ovale [48].

PDT

PDT is a non-thermal treatment modality that utilizes a laser-activated photosensitizing dye, specifically vitexoporphin, to induce vascular closure, tumor necrosis, and apoptosis. This approach is applicable in the management of small choroidal melanomas [49]. Nevertheless, the presence of tumor pigmentation can

diminish the efficacy of PDT, thereby limiting its application primarily to small, unpigmented choroidal melanomas. Additionally, PDT serves as an adjuvant to radiation therapy and as a complementary treatment following the failure of radiation therapy.

Particle radiation therapy

In contrast to traditional photon radiation therapy, particle radiation therapy is a form of long-distance radiation treatment that employs charged particles, such as protons, carbon ions, and helium ions, as the radiation source. These particle beams are accelerated to high energies using an accelerator and are delivered into the body with precision, concentrating their energy release at the target site. Upon acceleration, the particle beam is precisely directed into the body, where it releases energy at the lesion site, resulting in a sharply attenuated energy distribution known as the Bragg peak. Consequently, particle radiation therapy

Local excision of tumor

Initially, local excision of tumors was employed for the management of residual tumors following radiation therapy. Subsequently, it has been reported as the preferred treatment modality for UM [52]. This surgical procedure is categorized into two types: external resection, which involves the removal of the entire tumor through a scleral incision, and internal resection, which entails the removal of the entire tumor via the vitreous. External resection is typically indicated for melanomas of the iris, ciliary body, and peripheral choroid, whereas internal resection is indicated for choroidal melanomas situated posterior to the equator. The local excision of the tumor facilitates the acquisition of fresh tissue specimens for histopathological examination and genetic analysis, while also preserving ocular integrity and vision. In cases where tumor remnants are present on the scleral surface or the tumor is in proximity to the surgical margins, local excision may be

Eyeball removal

Enucleation is indicated for large and advanced uveal melanoma (UM), characterized by a tumor base diameter exceeding 20 mm or a thickness greater than 12 mm, as

Orbital content removal

In instances where UM extends into the orbit, an orbital exenteration procedure is performed, with efforts made to

Treatment for distant metastasis of UM

Currently, there is no definitive and effective treatment for distant metastasis of UM. For isolated metastatic lesions in

minimizes damage to surrounding healthy tissues, offers precise targeting, and ensures uniform dose distribution, making it the preferred method for UM radiation therapy [46, 50]. The study results indicate that particle radiation therapy provides superior tumor control compared to photon radiation therapy, although it may be associated with a higher likelihood of anterior foregut complications [51]. Due to its precise targeting capabilities, particle radiation therapy is highly versatile and particularly effective for treating challenging tumors located near the macula or optic disc.

augmented with adjunctive compressive radiation therapy to mitigate the risk of recurrence. Although the outcomes of local tumor excision are generally favorable, the procedure is technically demanding and necessitates a high level of surgical expertise and skill.

The established surgical criteria for the local excision of uveal melanoma (UM) include [53]: (1) a maximum tumor base diameter of ≤ 15 mm; (2) absence of local infiltration, with no involvement of the sclera or orbit; and (3) absence of systemic metastasis. Conversely, contraindications for surgical intervention include [53]: (1) extraocular infiltration or distant metastasis; (2) systemic conditions precluding surgical tolerance; and (3) the presence of flat, diffuse-shaped tumors, among others. The primary complications associated with the local excision of tumors encompass retinal detachment, proliferative vitreoretinopathy, and hemorrhage.

well as in cases involving optic nerve invasion, orbital extension, or secondary glaucoma [54].

preserve the eyelids to promote expedited postoperative recovery.

the liver, local interventions such as perfusion, embolization, and ablation may be considered, alongside

other recommended approaches including immunotherapy, cytotoxic regimens, and targeted therapy [55]. For patients

with confirmed distant metastases, participation in clinical trials is advised when feasible.

Recommendations for UM therapy

Treatment for UM is tailored to the individual. Radiation therapy is the most commonly employed method for managing UM, particularly for small to medium-sized tumors. In cases of large tumors, enucleation is the prevalent treatment option. For patients with specific indications, local excision of the tumor, TTT, and particle

radiation therapy can also yield favorable therapeutic outcomes. UM is characterized by mutations in key oncogenes, abnormalities in signaling pathways, and epigenetic imbalances. Research targeting the pathogenesis of UM may offer new opportunities for the development of molecularly targeted therapies.

HISTOPATHOLOGICAL DIAGNOSIS

Histopathologic features

UMs exhibit considerable variability in their gross morphology, presenting as dome-shaped, mushroom-shaped, or flat and diffuse lesions. The tumors may be pigmented, non-pigmented, or a combination of both. Choroidal melanomas that breach Bruch's membrane and assume a mushroom shape are prone to extrusion through this membrane, leading to vasodilatation and congestion within the tumor vasculature. Extraocular dissemination of UM typically occurs via the vortex veins or through the nerves and blood vessels traversing the sclera. Choroidal melanomas predominantly exhibit vertical growth and may penetrate Bruch's membrane, extending into the eye, potentially crossing the retina into the vitreous body, or spreading through the scleral channels or directly infiltrating the sclera, thereby extruding and

infiltrating in both inward and outward directions.

The histopathological classification of UMs is divided into four categories: spindle-cell, epithelioid cellular, mixed-cellular, and necrotic types. Melanomas composed of at least 90% spindle cells are classified as spindle cell melanomas, while those predominantly consisting of at least 90% epithelioid cells are designated as epithelioid cell melanomas. Tumors containing at least 10% epithelioid cells and up to 90% spindle cells are categorized as mixed-cell melanomas, which constitute the most common subtype. The primary immunohistochemical markers employed in the identification of UM include S-100 protein, anti-melanoma specific antibody (HMB-45), Melan-A, SRY-box transcription factor 10 (SOX10), and Vimentin [56, 57].

Histopathologically

Prognostic factors relevant to the prediction of UM outcomes include tumor cell type, mitotic activity, the mean diameter of the nucleoli of the 10 largest tumor cells,

microvessel density, extravascular stroma pattern, and the number of tumor-infiltrating lymphocytes and macrophages [58] (Figure 3).

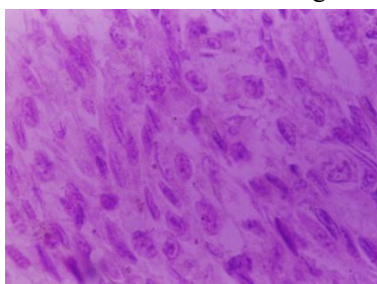


Figure 3: Histopathology of Choroidal Melanoma (HE×400). The tumor cells are spindle-shaped, with nuclear membrane folds forming striations in the center of the nucleus.

Table 4. Metastasis Risk Levels and Characteristics of Uveal Melanoma.

Risk Level	Detected Genes	Primary Tumor Characteristics (T) in TNM Staging	Tumor Histological Type	Follow-up Recommendations
Low Risk	Disomy of chromosome 3, gain of chromosome 6p, EIF1AX gene mutation	T1	Spindle cell type	Monitor every 12 months
Intermediate Risk	Disomy of chromosome 3, SF3B1 gene mutation	T2 and T3	Mixed cell type	Monitor every 6 to 12 months within 10 years
High Risk	Monosomy of chromosome 3, gain of chromosome 8q, BAP1 gene mutation, PRAME gene mutation	T4	Epithelioid cell type	Monitor every 3 to 6 months in the first 5 years, then every 6 to 12 months in the following 5 years

FOLLOW-UP AND PROGNOSIS

The management of UM necessitates regular monitoring of both ocular and systemic health. Post-radiation therapy, emphasis should be placed on evaluating the tumor and its associated complications, alongside the patient's overall health. In contrast, following enucleation, the primary focus should be on the patient's general condition.

A thorough ophthalmologic assessment is essential during follow-up visits, encompassing evaluations of visual acuity, intraocular pressure, and fundus condition, the latter being assessed after pupil dilation. Additionally, color photography and ultrasonography are indispensable for evaluating local tumor control. Techniques such as ultra-wide angle fundus imaging and fluorescein fundus angiography (FFA) are valuable for assessing peripheral fundus tumors and retinal vascular perfusion post-radiation therapy. Tumor recurrence predominantly occurs at the tumor margins, with central tumor recurrence and extraocular spread being less frequent.

Following the administration of radiation therapy, it is imperative to conduct follow-up examinations every 3 to 6 months during the initial two years, and subsequently every 6 to 12 months, to monitor for tumor recurrence and other potential complications. When available, genetic testing can be utilized to classify the metastatic risk grade, which, when integrated with the clinicopathological characteristics of the tumor, informs the postoperative follow-up strategy for patients (Table 4). The 5-year recurrence rate for small

UM tumors following palliative radiation therapy is 6%, and the 10-year recurrence rate is 11%. In contrast, the 5-year recurrence rate for large UM tumors is 13%. It is crucial to differentiate between tumor recurrence and inadequate tumor regression, the latter indicating nonresponsiveness to treatment. Recurrence is associated with an elevated risk of metastasis development [55].

The tumor's volume and its proximity to the optic disc and macula are critical determinants of the patient's visual prognosis. The primary factors contributing to the effects of PBRT on patients' vision include radiation retinopathy, radiation optic neuropathy, and cataracts. Although the peak incidence of radiation retinopathy occurs within five years post-radiation therapy, approximately 7% of patients develop related lesions 7-10 years following treatment [59]. Furthermore, about 69% of patients experience vision loss within a decade post-surgery, necessitating long-term monitoring for radiation retinopathy. Concurrently, periocular or intravitreal administration of tretinoin or anti-vascular endothelial growth factor (anti-VEGF) agents, and/or panretinal photocoagulation, may be employed to manage the condition [60].

Patients diagnosed with UM require comprehensive monitoring to facilitate the early detection of metastatic lesions. This monitoring predominantly involves liver-specific imaging, abdominal ultrasonography, and enhanced MRI, as well as enhanced CT scans of the chest,

abdomen, and pelvis. Whole-body PET-CT may also be employed when necessary. Additionally, it is essential to provide thorough psychological evaluation, counseling, and support to assist patients in managing ocular or

systemic discomfort, visual disturbances, alterations in appearance, and negative emotions such as worry, anxiety, and frustration regarding their future health, which may arise during treatment and follow-up.

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REFERENCES

1. Kaliki S, Shields CL, and Shields JA (2015) Uveal melanoma: estimating prognosis. *Journa* 63:93-102. http://doi: 10.4103/jmh.JMH_41_17
2. Shields CL, Kaliki S, Furuta M, Mashayekhi A, and Shields JA (2012) Clinical spectrum and prognosis of uveal melanoma based on age at presentation in 8,033 cases. *Journa* 32:1363-1372. <http://doi.org/10.3109/09286586.2010.498659>
3. Naseripoor M, Azimi F, Mirshahi R, Khakpoor G, Poorhosseingholi A, and Chaibakhsh S (2022) Global Incidence and Trend of Uveal Melanoma from 1943-2015: A Meta-Analysis. *Journa* 23:1791-1801. <http://doi.org/10.1080/09286580802521317>
4. Shields CL, Furuta M, Thangappan A, Nagori S, Mashayekhi A, Lally DR, et al. (2009) Metastasis of uveal melanoma millimeter-by-millimeter in 8033 consecutive eyes. *Journa* 127:989-998. <http://doi.org/10.1111/cxo.12210>
5. Kujala E, Mäkitie T, and Kivelä T (2003) Very long-term prognosis of patients with malignant uveal melanoma. *Journa* 44:4651-4659. <http://doi.org/10.1167/jovs.11-9228>
6. Yue H, Qian J, Yuan Y, Zhang R, Bi Y, Meng F, et al. (2017) Clinicopathological Characteristics and Prognosis for Survival after Enucleation of Uveal Melanoma in Chinese Patients: Long-term Follow-up. *Journa* 42:759-765. <http://doi.org/10.1038/s41598-018-36181-x>
7. Virgili G, Gatta G, Ciccolallo L, Capocaccia R, Biggeri A, Crocetti E, et al. (2007) Incidence of uveal melanoma in Europe. *Journa* 114:2309-2315. <http://doi.org/10.1016/j.opht.2016.12.011>
8. Liu YM, Li Y, Wei WB, Xu X, and Jonas JB (2015) Clinical Characteristics of 582 Patients with Uveal Melanoma in China. *Journa* 10:e0144562. <http://doi.org/10.1016/j.opht.2008.06.022>
9. Garg G, Finger PT, Kivelä TT, Simpson ER, Gallie BL, Saakyan S, et al. (2022) Patients presenting with metastases: stage IV uveal melanoma, an international study. *Journa* 106:510-517. <https://pubmed.ncbi.nlm.nih.gov/10860804>
10. Rietschel P, Panageas KS, Hanlon C, Patel A, Abramson DH, and Chapman PB (2005) Variates of survival in

Guidance Statement

All the experts involved in the development of this guideline declare that they adhere to an objective position, based on professional knowledge, research data and clinical experience, and that this guideline is formed after full discussion and unanimous agreement of all the experts.

Disclaimer

The contents of this guideline represent only the guidance of the experts involved in the development of this guideline for the reference of clinicians. Despite extensive consultation and discussion among experts, there are incomplete points. The recommendations provided in this guideline are not mandatory, and practices that are inconsistent with this guideline do not imply error or inappropriateness. There are still many issues to be explored in clinical practice, and ongoing and future clinical trials will provide further evidence. With the accumulation of clinical experience and the emergence of new treatments, this guideline will need to be revised and updated periodically in the future to bring more clinical benefits to the subjects.

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CONFLICTS OF INTEREST

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metastatic uveal melanoma. *Journa* 23:8076-8080.

<http://doi.org/10.1016/j.pscychresns.2015.01.018>

11. Finger PT, Kurli M, Reddy S, Tena LB, and Pavlick AC (2005) Whole body PET/CT for initial staging of choroidal melanoma. *Journa* 89:1270-1274.

<http://doi.org/10.12659/msm.897837>

12. Buder K, Gesierich A, Gelbrich G, and Goebeler M (2013) Systemic treatment of metastatic uveal melanoma: review of literature and future perspectives. *Journa* 2:674-686. <http://doi.org/10.1097/wnr.0000000000000471>

13. Sipos E, Hegyi K, Treszl A, Steiber Z, Mehes G, Dobos N, et al. (2017) Concurrence of chromosome 3 and 4 aberrations in human uveal melanoma. *Journa* 37:1927-1934. <http://doi.org/10.3892/mmr.2017.7006>

14. Dogrusöz M and Jager MJ (2018) Genetic prognostication in uveal melanoma. *Journa* 96:331-347. <http://doi: 10.1167/tvst.8.1.1>

15. Rodrigues M, Ait Rais K, Salviat F, Algret N, Simaga F, Barnhill R, et al. (2020) Association of Partial Chromosome 3 Deletion in Uveal Melanomas With Metastasis-Free Survival. *Journa* 138:182-188. <http://doi.org/10.1097/md.0000000000006139>

16. Carvajal RD, Sacco JJ, Jager MJ, Eschelman DJ, Olofsson Bagge R, Harbour JW, et al. (2023) Advances in the clinical management of uveal melanoma. *Journa* 20:99-115. <https://pubmed.ncbi.nlm.nih.gov/27045330>

17. He M, Chaurushiya MS, Webster JD, Kummerfeld S, Reja R, Chaudhuri S, et al. (2019) Intrinsic apoptosis shapes the tumor spectrum linked to inactivation of the deubiquitinase BAP1. *Journa* 364:283-285. <http://doi.org/10.7554/eLife.06356>

18. Harbour JW, Roberson ED, Anbunathan H, Onken MD, Worley LA, and Bowcock AM (2013) Recurrent mutations at codon 625 of the splicing factor SF3B1 in uveal melanoma. *Journa* 45:133-135. <http://doi.org/10.1371/journal.pone.0024124>

19. Li Y, Shi J, Yang J, Ge S, Zhang J, Jia R, et al. (2020) Uveal melanoma: progress in molecular biology and therapeutics. *Journa* 12:1758835920965852. <http://doi.org/10.1016/j.nicl.2015.09.018>

20. Smit KN, Jager MJ, de Klein A, and Kiliç E (2020) Uveal melanoma: Towards a molecular understanding. *Journa* 75:100800. <https://doi.org/10.1007/s00347-005-1299-y>

21. Fuentes-Rodriguez A, Mitchell A, Guérin SL, and Landreville S (2024) Recent Advances in Molecular and Genetic Research on Uveal Melanoma. *Journa* 13.

<http://doi.org/10.1111/j.1600-0447.1983.tb09716.x>

22. Li Y, Jia R, and Ge S (2017) Role of Epigenetics in Uveal Melanoma. *Journa* 13:426-433. <http://doi: 10.1016/j.ajo.2018.01.004>

23. Fratta E, Sigalotti L, Covre A, Parisi G, Coral S, and Maio M (2013) Epigenetics of melanoma: implications for immune-based therapies. *Journa* 5:1103-1116. <http://doi:10.1016/j.fertnstert.2016.09.046>

24. Chai P, Yu J, Jia R, Wen X, Ding T, Zhang X, et al. (2020) Generation of onco-enhancer enhances chromosomal remodeling and accelerates tumorigenesis. *Journa* 48:12135-12150. <http://doi: 10.1136/bjo.84.1.76>. PMID: 10611104

25. Yu J, Chai P, Xie M, Ge S, Ruan J, Fan X, et al. (2021) Histone lactylation drives oncogenesis by facilitating m(6)A reader protein YTHDF2 expression in ocular melanoma. *Journa* 22:85. <http://doi: 10.1001/jama.286.17.2114>

26. Jia R, Chai P, Wang S, Sun B, Xu Y, Yang Y, et al. (2019) m(6)A modification suppresses ocular melanoma through modulating HINT2 mRNA translation. *Journa* 18:161. <http://doi: 10.1016/j.psyneuen.2015.04.022>

27. Xing Y, Wen X, Ding X, Fan J, Chai P, Jia R, et al. (2017) CANT1 lncRNA Triggers Efficient Therapeutic Efficacy by Correcting Aberrant Incing Cascade in Malignant Uveal Melanoma. *Journa* 25:1209-1221. <http://doi.org/10.1093/brain/122.9.1781>

28. Chai P, Jia R, Li Y, Zhou C, Gu X, Yang L, et al. (2022) Regulation of epigenetic homeostasis in uveal melanoma and retinoblastoma. *Journa* 89:101030. <http://doi.org/10.3233/rnn-120267>

29. Shields CL, Sioufi K, Robbins JS, Barna LE, Harley MR, Lally SE, et al. (2018) LARGE UVEAL MELANOMA (≥10 MM THICKNESS): Clinical Features and Millimeter-by-Millimeter Risk of Metastasis in 1311 Cases. The 2018 Albert E. Finley Lecture. *Journa* 38:2010-2022. <http://doi.org/10.1212/WNL.0b013e31821a44c1>

30. Hood CT, Schoenfield LR, Torres V, and Singh AD (2011) Iris melanoma. *Journa* 118:221-222. <http://doi.org/10.1016/j.neuropsychologia.2004.04.016>

31. LoRusso FJ, Boniuk M, and Font RL (2000) Melanocytoma (magnocellular nevus) of the ciliary body: report of 10 cases and review of the literature. *Journa* 107:795-800. <http://doi.org/10.1093/brain/awq210>

32. Li X, Wang L, Zhang L, Tang F, and Wei X (2020) Application of Multimodal and Molecular Imaging Techniques

in the Detection of Choroidal Melanomas. *Journa* 10:617868.
<http://doi.org/10.1007/s00429-017-1406-2>

33. Conway RM, Chew T, Golchet P, Desai K, Lin S, and O'Brien J (2005) Ultrasound biomicroscopy: role in diagnosis and management in 130 consecutive patients evaluated for anterior segment tumours. *Journa* 89:950-955.
<http://doi.org/10.1038/npp.2009.131>

34. Yang WL, Wei WB, and Li DJ (2013) [Characteristics of choroidal melanoma in contrast-enhanced ultrasonography]. *Journa* 49:428-432.
<http://doi.org/10.3389/fnana.2011.00040>

35. Ferreira TA, Grech Fonk L, Jaarsma-Coes MG, van Haren GGR, Marinkovic M, and Beenakker JM (2019) MRI of Uveal Melanoma. *Journa* 11.
<http://doi.org/10.1016/j.neubiorev.2007.07.010>

36. Solnik M, Paduszyńska N, Czarnecka AM, Synoradzki KJ, Yousef YA, Chorągiewicz T, et al. (2022) Imaging of Uveal Melanoma-Current Standard and Methods in Development. *Journa* 14.
[http://doi.org/10.1002/1097-0029\(20001001\)51:1<85::Aid-je mt9>3.0.Co;2-0](http://doi.org/10.1002/1097-0029(20001001)51:1<85::Aid-je mt9>3.0.Co;2-0)

37. Obuchowska I and Konopińska J (2022) Importance of Optical Coherence Tomography and Optical Coherence Tomography Angiography in the Imaging and Differentiation of Choroidal Melanoma: A Review. *Journa* 14.
<http://doi.org/10.1016/j.pneurobio.2013.02.003>

38. Shields CL, Say EA, Samara WA, Khoo CT, Mashayekhi A, and Shields JA (2016) OPTICAL COHERENCE TOMOGRAPHY ANGIOGRAPHY OF THE MACULA AFTER PLAQUE RADIOTHERAPY OF CHOROIDAL MELANOMA: Comparison of Irradiated Versus Nonirradiated Eyes in 65 Patients. *Journa* 36:1493-1505.
<http://doi.org/10.1212/01.wnl.0000038905.75660.bd>

39. Hui KH, Pfeiffer ML, and Esmaeli B (2012) Value of positron emission tomography/computed tomography in diagnosis and staging of primary ocular and orbital tumors. *Journa* 26:365-371.
<http://doi.org/10.1016/j.neuron.2015.06.005>

40. Frizziero L, Midena E, Trainiti S, Londei D, Bonaldi L, Bini S, et al. (2019) Uveal Melanoma Biopsy: A Review. *Journa* 11.
<http://doi.org/10.1016/j.ejpain.2004.11.001>

41. Cao SS, Li HY, Xu QG, Tan SY, and Wei SH (2016) [Clinical features of neurosyphilis with optic neuritis as an initial finding]. *Journa* 52:898-904.
<http://doi.org/10.1016/j.pain.2008.02.034>

42. Tretiakow D, Mikaszewski B, and Skorek A (2020)

The role of fine-needle aspiration biopsy (FNAB) in the diagnostic management of parotid gland masses with emphasis on potential pitfalls. *Journa* 277:2939-2940.
<http://doi.org/10.1212/01.wnl.0000038905.75660.bd>

43. Singh AD and Kivelä T (2005) The collaborative ocular melanoma study. *Journa* 18:129-142, ix.
<http://doi.org/10.1016/j.neuron.2015.06.005>

44. Amin MB, Edge SB, Greene FL, Byrd DR, Brookland RK, Washington MK, et al., AJCC cancer staging manual. Vol. 1024. 2017: Springer.
<http://doi.org/10.1016/j.ejpain.2004.11.001>

45. Weis E, Surgeoner B, Salopek TG, Cheng T, Hyrcza M, Kostaras X, et al. (2023) Management of Uveal Melanoma: Updated Cancer Care Alberta Clinical Practice Guideline. *Journa* 31:24-41.
<http://doi.org/10.1016/j.pain.2008.02.034>

46. Network NCC (2008) NCCN clinical practice guidelines in oncology. *Journa*.
<http://doi.org/10.1097/md.0000000000006139>

47. Diener-West M, Earle JD, Fine SL, Hawkins BS, Moy CS, Reynolds SM, et al. (2001) The COMS randomized trial of iodine 125 brachytherapy for choroidal melanoma, III: initial mortality findings. COMS Report No. 18. *Journa* 119:969-982.
<https://pubmed.ncbi.nlm.nih.gov/27045330>

48. Aaberg TM, Jr., Bergstrom CS, Hickner ZJ, and Lynn MJ (2008) Long-term results of primary transpupillary thermal therapy for the treatment of choroidal malignant melanoma. *Journa* 92:741-746.
<http://doi.org/10.7554/eLife.06356>

49. Yordi S, Soto H, Bowen RC, and Singh AD (2021) Photodynamic therapy for choroidal melanoma: What is the response rate? *Journa* 66:552-559.
<http://doi.org/10.1371/journal.pone.0024124>

50. Rao PK, Barker C, Coit DG, Joseph RW, Materin M, Rengan R, et al. (2020) NCCN Guidelines Insights: Uveal Melanoma, Version 1.2019. *Journa* 18:120-131.
<http://doi.org/10.1016/j.nicl.2015.09.018>

51. Wang Z, Nabhan M, Schild SE, Stafford SL, Petersen IA, Foote RL, et al. (2013) Charged particle radiation therapy for uveal melanoma: a systematic review and meta-analysis. *Journa* 86:18-26.
<https://doi.org/10.1007/s00347-005-1299-y>

52. Damato BE (2011) Local resection of uveal melanoma. *Journa* 49:66-80.
<http://doi.org/10.1111/j.1600-0447.1983.tb09716.x>

53. Hamza HS and Elhusseiny AM (2018) Choroidal Melanoma Resection. *Journa* 25:65-70.
<http://doi:10.1016/j.ajo.2018.01.004>

54. Amaro TA, Yazigi L, and Erwenne C (2010) Depression and quality of life during treatment of ocular bulb removal in individuals with uveal melanoma. *Journa* 19:476-481.
<http://doi:10.1016/j.fertnstert.2016.09.046>
55. Jager MJ, Shields CL, Cebulla CM, Abdel-Rahman MH, Grossniklaus HE, Stern MH, et al. (2020) Uveal melanoma. *Journa* 6:24.
<http://doi.org/10.3233/rnn-120267>
56. Saraiva VS, Caissie AL, Segal L, Edelstein C, and Burnier MN, Jr. (2005) Immunohistochemical expression of phospho-Akt in uveal melanoma. *Journa* 15:245-250.
<http://doi.org/10.1212/WNL.0b013e31821a44c1>
57. Fernandes BF, Odashiro AN, Saraiva VS, Logan P, Anteck E, and Burnier MN, Jr. (2007) Immunohistochemical expression of melan-A and tyrosinase in uveal melanoma. *Journa* 6:6.
<http://doi.org/10.1016/j.neuropsychologia.2004.04.016>
58. Berus T, Halon A, Markiewicz A, Orlowska-Heitzman J, Romanowska-Dixon B, and Donizy P (2017) Clinical, Histopathological and Cytogenetic Prognosticators in Uveal Melanoma - A Comprehensive Review. *Journa* 37:6541-6549.
<http://doi.org/10.1093/brain/awq210>
59. Wisely CE, Hadziahmetovic M, Reem RE, Hade EM, Nag S, Davidorf FH, et al. (2016) Long-term visual acuity outcomes in patients with uveal melanoma treated with 125I episcleral OSU-Nag plaque brachytherapy. *Journa* 15:12-22.
<http://doi.org/10.1007/s00429-017-1406-2>
60. Nhàn NTT, Ganesh S, Maidana DE, Heiferman MJ, and Yamada KH (2025) Uveal melanoma with a GNA11/GNAQ mutation secretes VEGF for systemic spread. *Journa* 10:51.
<http://doi.org/10.1038/npp.2009.131>